

# Cooperative C3- and C13-Substituent Effects on Synthetic Chlorophyll Derivatives

Shin-ichi Sasaki,<sup>[a,b]</sup> Mari Yoshizato,<sup>[a]</sup> Michio Kunieda,<sup>[a]</sup> and Hitoshi Tamiaki\*<sup>[a]</sup>

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C3-vinyl and C13-keto-carbonyl groups of methyl pyropheophorbide-*a*, a chlorophyll-*a* derivative, were systematically modified to investigate the substituent effects along the  $y$ -axis of chlorin macrocycles. C13<sup>1</sup> deoxygenation resulted in blueshifts of the  $Q_y$  absorption maxima and a large reduction of their redox potentials, whereas introduction of the dicya-

nomethylene group at the 13<sup>1</sup>-position caused the opposite effect on both optical and electrochemical properties. Among the nine chlorins examined, the  $Q_y$  peak positions could be varied from 647 to 736 nm by the cooperative effects of the diagonal substituents' combinations.

## Introduction

Chlorophyll(Chl)-*a* (Figure 1, a) possessing the C3-vinyl group is a representative dye molecule in natural photosynthesis, and its C3-formyl analogue, Chl-*d*, is observed in some cyanobacteria.<sup>[1]</sup> This difference between the C3-vinyl and formyl groups causes a shift of the  $Q_y$  peak positions: 661 (Chl-*a*) and 686 nm (Chl-*d*) in ethyl ether.<sup>[2]</sup> On the other hand, bacteriochlorophyll-*d* has the C3-(1-hydroxyethyl) group on its chlorin macrocycle to give a blueshifted  $Q_y$  maximum at 650 nm as its monomeric form in ethyl ether.<sup>[3]</sup> To clarify such C3-substituent effects on optical properties, we reported a series of Chl derivatives possessing different C3-substituents.<sup>[4,5]</sup>

Since alteration of the C3-functional group brings visual color changes in solution and drastic  $Q_y$  peak shifts, introduction of a molecular recognition site at the 3-position successfully led to the development of chlorophyll-based chemosensors (Figure 1, b).<sup>[6]</sup> One of the advantages in using the chlorin unit for the design of chromo/fluororeceptors is its intense  $Q_y$  band, which is located at the longer-wavelength region. Because this characteristic band is dependent on the transition moment along the  $y$ -axis, transformation of the C13-functional group would also be useful for tuning the optical properties of the red-most bands. Thus spectral characteristics of C3-functionalized chlorins are expected to be effectively controlled by the simultaneous substituent effects at the diagonal 13-position.

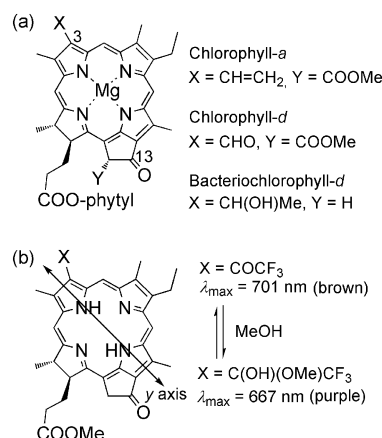


Figure 1. Chemical structures of (a) some natural (bacterio)chlorophylls and (b) an example of a chlorophyll-based chemosensor.

In this study we synthesized C3-substituted chlorins possessing a dicyanomethylene group at the 13<sup>1</sup>-position or lacking any C13-functional group and compared their physical properties with those of the corresponding C13<sup>1</sup>-oxochlorins. The cooperative effects of C3/C13-functional groups were elucidated based on absorption and emission spectroscopy together with cyclic voltammetry.

## Results and Discussion

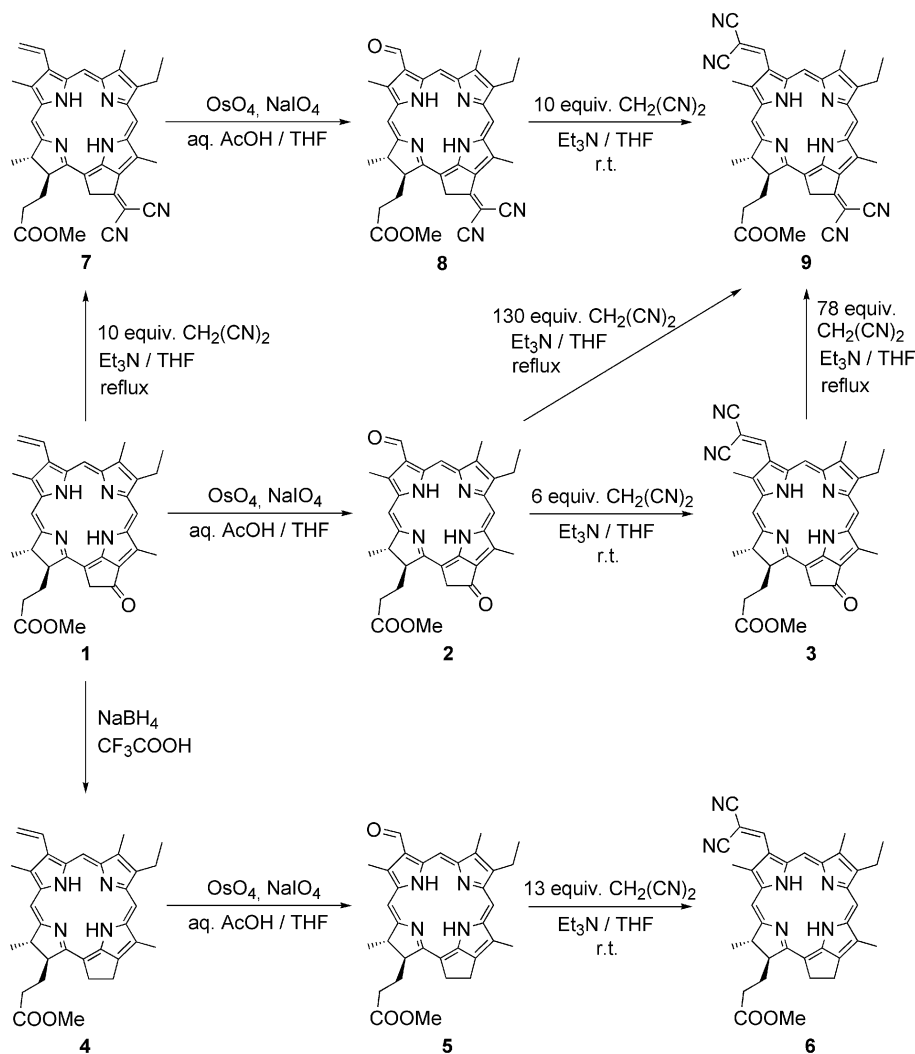
### Synthesis of Chlorophyll Derivatives 1–9

Chl-*a* was extracted from *Spirulina geitleri* and converted to methyl pyropheophorbide-*a* (**1**)<sup>[7]</sup> and its C13<sup>1</sup>-deoxo analogue **4**<sup>[8]</sup> according to the literature procedure. Oxidative cleavage of the C3-vinyl to formyl group (**1**  $\rightarrow$  **2**,<sup>[7]</sup> **4**  $\rightarrow$  **5**)<sup>[9]</sup>; Scheme 1) and the successive Knoevenagel reaction of the 3-formylchlorins with malononitrile (**2**  $\rightarrow$  **3**, **5**  $\rightarrow$  **6**) were

[a] Department of Bioscience and Biotechnology, Faculty of Science and Engineering, Ritsumeikan University, Kusatsu, Shiga 525-8577, Japan  
 Fax: +81-77-561-2659  
 E-mail: tamiaki@se.ritsumei.ac.jp

[b] Nagahama Institute of Bio-Science and Technology, Nagahama, Shiga 526-0829, Japan

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Scheme 1. Synthesis of chlorophyll derivatives 1–9.

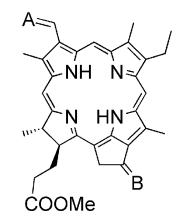
carried out by using previously reported procedures.<sup>[4]</sup> On the other hand, the Knoevenagel reaction was found to be applicable for the introduction of a dicyanomethylene group at the 13<sup>1</sup>-position. When the C13-keto-carbonyl group on **1** was treated with excess malononitrile in the presence of triethylamine in refluxing THF, the corresponding C13<sup>1</sup>-dicyanomethylene-chlorin **7** was obtained in 67% yield. The following oxidative cleavage of the C3-vinyl group of **7** and subsequent Knoevenagel reaction proceeded smoothly to afford **8** (70%) and **9** (41%) with less interference of the C13<sup>1</sup>-dicyanomethylene group. Alternatively, one-pot reaction of the 3- and 13-positions of **2** with malononitrile afforded **9** via **3**, as its counterpart **8** was not isolated even under mild conditions: its stepwise synthesis from **2** slightly improved the yield of **9** (**2**  $\rightarrow$  **9**: 7%, **2**  $\rightarrow$  **3**  $\rightarrow$  **9**:  $88 \times 12\% = 11\%$ ).

### Optical Properties of Synthetic Chlorins 1–9

The electronic absorption and emission spectra of 3-vinyl-, 3-formyl-, and 3-(dicyanoethenyl)chlorins are shown in

Figure 2, and their absorption maxima and fluorescence data are summarized in Table 1. When the C13-carbonyl group of chlorin derivatives is reduced to the C13-deoxo moiety, the  $Q_y$  peak maxima were shifted to a lower wavelength with a decrease of the relative peak absorbances ( $A_{\text{rel}}$ ). On the contrary, replacement of the C13-carbonyl with the dicyanoethenyl group caused redshifts of the  $Q_y$  peak. The observed  $Q_y$  peak positions are apparently related to the electron-withdrawing ability of the C13-functional group [ $B = \text{H}_2 \rightarrow \text{O} \rightarrow \text{C}(\text{CN})_2$  in Table 1], which is similar to the C3-substituent effects previously reported.<sup>[5]</sup> In view of the C13-substituent effect, the  $Q_y$  peak shift ( $940 \text{ cm}^{-1}$ ) accompanied by the conversion of the C3-vinyl to dicyanoethenyl through the formyl group of C13<sup>1</sup>-deoxochlorins **4**  $\rightarrow$  **6** ( $B = \text{H}_2$ ,  $\lambda_{\text{abs}} = 647 \rightarrow 689 \text{ nm}$ ) is larger than that of C13<sup>1</sup>-oxo-chlorins **1**  $\rightarrow$  **3** ( $B = \text{O}$ ,  $\lambda_{\text{abs}} = 667 \rightarrow 706 \text{ nm}$ ,  $830 \text{ cm}^{-1}$ ), whereas 13<sup>1</sup>-(dicyanomethylene)chlorins **7**  $\rightarrow$  **9** showed the least effect [ $B = \text{C}(\text{CN})_2$ ,  $\lambda_{\text{abs}} = 704 \rightarrow 736 \text{ nm}$ ,  $620 \text{ cm}^{-1}$ ]. An electron-withdrawing group seems to decrease the substituent effects at the diagonal position. Similar cooperative effects were also observed from changes

Table 1. Visible absorption ( $\lambda_{\text{abs}}$ ), fluorescence emission maxima ( $\lambda_{\text{em}}$ ), and fluorescence quantum yields ( $\Phi$ ) of chlorophyll derivatives **1–9** in  $\text{CH}_2\text{Cl}_2$ .



Compound	A	B	Absorption, $\lambda_{\text{abs}}$ ( $A_{\text{rel}}^{[a]}$ ) [nm] [fwhm] <sup>[b]</sup> [cm <sup>-1</sup> ]		Fluorescence emission <sup>[c]</sup>		
			$Q_x$	$Q_y$	$\lambda_{\text{em}}$ [nm]	$\Delta$ [cm <sup>-1</sup> ] <sup>[d]</sup>	$\Phi$ [%]
<b>1</b>	CH <sub>2</sub>	O	509, 539	609, 667 (0.44) [473]	672	110	22
<b>2</b>	O	O	521, 554	632, 694 (0.81) [435]	699	100	18
<b>3</b>	C(CN) <sub>2</sub>	O	522, 568	643, 706 (0.55) [803]	725	370	17
<b>4</b>	CH <sub>2</sub>	H <sub>2</sub>	501	592, 647 (0.24) [407]	650	70	17
<b>5</b>	O	H <sub>2</sub>	514, 551	610, 665 (0.34) [452]	667	50	14
<b>6</b>	C(CN) <sub>2</sub>	H <sub>2</sub>	580	636, 689 (0.34) [928]	715	530	10
<b>7</b>	CH <sub>2</sub>	C(CN) <sub>2</sub>	534, 574	643, 704 (1.10) [505]	712	160	25
<b>8</b>	O	C(CN) <sub>2</sub>	543, 584	663, 727 (1.30) [492]	733	110	21
<b>9</b>	C(CN) <sub>2</sub>	C(CN) <sub>2</sub>	545, 590	669, 736 (0.97) [802]	749	250	20

[a] Relative peak absorbance ( $A_{\text{rel}}$ ) was based on the most intense Soret peak. [b] Full width at half maximum. [c] Excited at Soret band. [d] Stokes shift ( $\Delta/\text{cm}^{-1}$ ) =  $[1/\lambda_{\text{abs}}(Q_y(0,0)) - 1/\lambda_{\text{em}}] \times 10^7$ .

of the C3-position; the  $Q_y$  peak shifts of **4** to **7** ( $A = \text{CH}_2$ ,  $\lambda_{\text{abs}} = 647 \rightarrow 704 \text{ nm}$ ,  $1250 \text{ cm}^{-1}$ ) and **5** to **8** ( $A = \text{O}$ ,  $\lambda_{\text{abs}} = 665 \rightarrow 727 \text{ nm}$ ,  $1280 \text{ cm}^{-1}$ ) are larger than that of 3-(di-cyanoethenyl)chlorins **6** to **9** [ $A = \text{C}(\text{CN})_2$ ,  $\lambda_{\text{abs}} = 689 \rightarrow 736 \text{ nm}$ ,  $930 \text{ cm}^{-1}$ ].

of the 3/13-positions of the group. This functional group seems to bring a large Stokes shift relative to other types of combinations of the substituents ( $A = \text{CH}_2$ ,  $\text{O}$ ;  $B = \text{H}_2$ ,  $\text{O}$ ). The fluorescence quantum yields were varied between 10 and 25%, and specific increase or quenching was not observed. Although the differences are not very large, electron-withdrawing groups at the 3-position decrease the  $\Phi$  values, whereas the C13-functional groups appear to have the opposite effect.

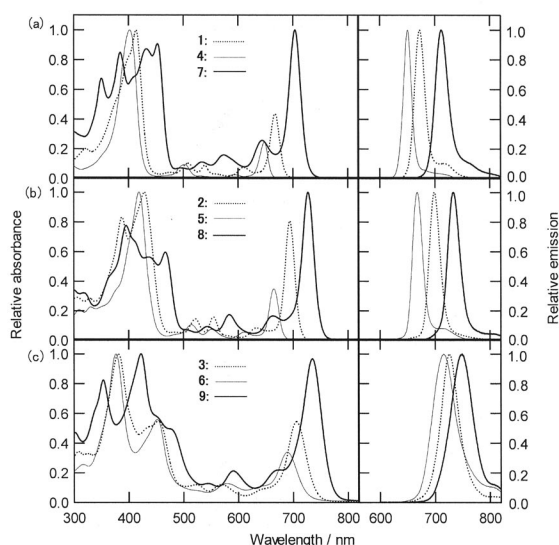


Figure 2. Electronic absorption (left) and fluorescence emission spectra (right) of (a) 3-vinylchlorins **1** (dotted line), **4** (solid thin line), and **7** (solid thick line), (b) 3-formylchlorins **2** (dotted line), **5** (solid thin line), and **8** (solid thick line), and (c) 3-(dicyanoethenyl)-chlorins **3** (dotted line), **6** (solid thin line), and **9** (solid thick line) in  $\text{CH}_2\text{Cl}_2$ . All spectra were normalized at their most intense maxima.

It should be noted that the dicyanomethylene group causes not only the redshifts of the  $Q_x/Q_y$  maxima but also characteristic peak broadening of the Soret band, regardless

### Electrochemical Properties of Synthetic Chlorins **1–9**

Redox potentials of chlorins **1–9** are summarized in Table 2.<sup>[10]</sup> Oxidation potentials ( $E_{1/2}^{\text{ox}}$ ) of three-type C3-substituted chlorins [ $A = \text{CH}_2$ ,  $\text{O}$ , or  $\text{C}(\text{CN})_2$ ] increased in the

Table 2. Redox potentials of chlorophyll derivatives **1–9**.<sup>[a]</sup>

	A <sup>[b]</sup>	B <sup>[b]</sup>	$E_{1/2}^{\text{ox}}$ [mV]	$E_{1/2}^{\text{red}}$ [mV]	$\Delta E$ [mV] <sup>[c]</sup>	$10^7/\lambda_{\text{abs}}$ [cm <sup>-1</sup> ] <sup>[d]</sup>
<b>1</b>	CH <sub>2</sub>	O	390	-1590	1980	15000
<b>2</b>	O	O	480	-1400	1880	14400
<b>3</b>	C(CN) <sub>2</sub>	O	500	— <sup>[e]</sup>	—	14200
<b>4</b>	CH <sub>2</sub>	H <sub>2</sub>	110	-1930	2040	15500
<b>5</b>	O	H <sub>2</sub>	240	-1720	1960	15000
<b>6</b>	C(CN) <sub>2</sub>	H <sub>2</sub>	280	— <sup>[e]</sup>	—	14500
<b>7</b>	CH <sub>2</sub>	C(CN) <sub>2</sub>	450	-1370	1820	14200
<b>8</b>	O	C(CN) <sub>2</sub>	540	-1260	1800	13800
<b>9</b>	C(CN) <sub>2</sub>	C(CN) <sub>2</sub>	580	— <sup>[e]</sup>	—	13600

[a] Cyclic voltammograms were recorded in  $\text{CH}_2\text{Cl}_2/0.1 \text{ M Bu}_4\text{N-ClO}_4$  with a scan rate of  $10 \text{ mV/s}$  at ca.  $1 \text{ mM}$  sample solutions. The redox potential values are reported based on the oxidation potential of ferrocene as a standard. [b] Refer to the structure in Table 1. [c] The difference between oxidation and reduction potentials:  $\Delta E = E_{1/2}^{\text{ox}} - E_{1/2}^{\text{red}}$ . [d] The wavenumber of the  $Q_y(0,0)$  peak maxima. [e] Peaks were not clearly observed.

order of  $B = \text{CH}_2 \rightarrow \text{O} \rightarrow \text{C}(\text{CN})_2$  and reduction potentials ( $E_{1/2}^{\text{red}}$ ) showed more positive values in this sequence. A similar tendency was observed in changes of  $A = \text{CH}_2 \rightarrow \text{O} \rightarrow \text{C}(\text{CN})_2$  for C13<sup>1</sup>-substituted chlorins [ $B = \text{O}$ ,  $\text{H}_2$ , or  $\text{C}(\text{CN})_2$ ]. Electron-withdrawing substituents at the 3/13-positions decrease the electron density on the chlorin macrocycle and make the release of one electron more difficult. As expected, an almost linear relationship (correlation coefficient  $r^2 = 0.966$ ) can be seen when the potential differences of  $\Delta E (= E_{1/2}^{\text{ox}} - E_{1/2}^{\text{red}})$  are plotted versus the energy level of  $Q_y$  maxima ( $10^7/\lambda_{\text{abs}}$ ) in wavenumber units (see Figure S5 in the Supporting Information).

## Conclusions

We have demonstrated the transformation of methyl pyropheophorbide-*a* to 13<sup>1</sup>-(dicyanomethylene)chlorin derivatives, which caused drastic spectral changes, especially in their  $Q_y$  bands. The modified dye molecules should be useful for the development of new chromo/fluororeceptors<sup>[6]</sup> and for application to dye-sensitized solar cells,<sup>[11]</sup> as the C3-formyl groups of **2**, **5**, and **8** could be further altered.<sup>[4–6,12]</sup>

## Experimental Section

**General Methods:** Electronic absorption and fluorescence emission spectra were measured on Hitachi U-3500 and F-4500 spectrometers, respectively. Fluorescence quantum yields in  $\text{CH}_2\text{Cl}_2$  were determined by using a Hamamatsu Photonics C9920-02 spectrometer. All melting points were measured with a Yanagimoto micro melting apparatus and were uncorrected. <sup>1</sup>H NMR spectra were measured with a JEOL JNM-600HR spectrometer; chemical shifts ( $\delta$ ) in  $\text{CDCl}_3$  are expressed in parts ppm relative to TMS ( $\delta = 0.00$  ppm) as an internal reference and peaks were assigned by <sup>1</sup>H-<sup>1</sup>H COSY and NOESY spectra. FTIR spectra were measured with a Shimadzu IRAffinity-1 spectrophotometer. Electrochemical measurements were carried out by a BAS CV-50W voltammetric analyzer with a conventional three-electrode system. FAB-MS were measured with a JEOL GCmate II spectrometer. FAB-MS samples were dissolved in  $\text{CH}_2\text{Cl}_2$  and *m*-nitrobenzyl alcohol and glycerol were used as the matrices and PEG600 was added as an internal reference.

Methyl pyropheophorbide-*a* (**1**),<sup>[7]</sup> methyl pyropheophorbide-*d* (**2**),<sup>[7]</sup> methyl 3<sup>2</sup>,3<sup>2</sup>-dicyano-pyropheophorbide-*a* (**3**),<sup>[4]</sup> methyl 13<sup>1</sup>-deoxo-pyropheophorbide-*a* (**4**),<sup>[8]</sup> and methyl 13<sup>1</sup>-deoxo-pyropheophorbide-*d* (**5**)<sup>[9]</sup> were prepared as previously reported. THF was distilled from  $\text{CaH}_2$  before use. Other solvents and reagents were employed as purchased without further purification. All synthetic procedures were carried out in the dark.

**Methyl 3<sup>2</sup>,3<sup>2</sup>-Dicyano-13<sup>1</sup>-deoxo-pyropheophorbide-*a* (**6**):** According to the reported procedure,<sup>[4]</sup> Knoevenagel reaction of 3-formylchlorin **5** (30 mg, 0.056 mmol) was performed by malononitrile (50 mg, 0.76 mmol) and  $\text{Et}_3\text{N}$  (24 mg, 0.24 mmol) in refluxing THF (20 mL) for 6 h. The crude product was purified by chromatography on silica gel ( $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$ , 1:99) followed by recrystallization from  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  to give **6** (16 mg, 48%) as brown crystals; m.p. 208–209 °C. UV/Vis ( $\text{CH}_2\text{Cl}_2$ ):  $\lambda_{\text{max}}$  ( $A_{\text{rel}}$ ) = 689 (0.34), 636 (0.10), 580 (0.13), 451 (0.56), 376 (1) nm; Flu ( $\text{CH}_2\text{Cl}_2$ ):  $\lambda_{\text{ex}}$  =

450 nm);  $\lambda_{\text{em}}$  = 715 nm;  $\Phi_{\text{Flu}}$  ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 450 nm) = 10%. IR (film):  $\tilde{\nu}$  = 2228 ( $\text{C}\equiv\text{N}$ ), 1744 ( $17^2\text{-C=O}$ )  $\text{cm}^{-1}$ . <sup>1</sup>H NMR ( $\text{CDCl}_3$ ):  $\delta$  = 9.54 (s, 1 H, 5-H), 9.52 (s, 1 H, 3-CH), 9.40 (s, 1 H, 10-H), 8.94 (s, 1 H, 20-H), 4.93–4.88, 4.81–4.76 (m, each 1 H, 13<sup>1</sup>-CH<sub>2</sub>), 4.60 (dq,  $J$  = 2, 7 Hz, 1 H, 18-H), 4.46–4.43 (br-d, 1 H, 17-H), 4.09–3.99 (m, 2 H, 13-CH<sub>2</sub>), 3.80 (q,  $J$  = 8 Hz, 2 H, 8-CH<sub>2</sub>), 3.64, 3.60 (s, each 3 H, 2-CH<sub>3</sub>, 17<sup>2</sup>-CO<sub>2</sub>CH<sub>3</sub>), 3.45 (s, 3 H, 12-CH<sub>3</sub>), 3.41 (s, 3 H, 7-CH<sub>3</sub>), 2.82–2.76, 2.66–2.60, 2.32–2.23 (m, 1 H, 1 H, 2 H, 17-CH<sub>2</sub>CH<sub>2</sub>), 1.83 (d,  $J$  = 7 Hz, 3 H, 18-CH<sub>3</sub>), 1.75 (t,  $J$  = 8 Hz, 3 H, 8<sup>1</sup>-CH<sub>3</sub>), –1.08, –2.46 (s, each 1 H,  $\text{NH}\times 2$ ) ppm. MS (FAB): calcd. for  $\text{C}_{36}\text{H}_{36}\text{N}_6\text{O}_2$  584.2900; found 584.2946 [ $\text{M}$ ]<sup>+</sup>.

**Methyl 13<sup>1</sup>-Deoxo-13<sup>1</sup>-(dicyanomethylene)pyropheophorbide-*a* (**7**):** A mixture of chlorin **1** (27 mg, 0.050 mmol), malononitrile (33 mg, 0.50 mmol), and  $\text{Et}_3\text{N}$  (35 mg, 0.35 mmol) in THF (20 mL) was refluxed for 1 h. The solution was cooled to room temperature, poured into 1% aqueous HCl, and extracted with  $\text{CH}_2\text{Cl}_2$ . The extract was washed with water, dried with anhydrous  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The crude product was purified by silica-gel chromatography ( $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$ , 1:99) followed by recrystallization from  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  to give **7** (20 mg, 67%) as brown crystals; m.p. 232–236 °C. UV/Vis ( $\text{CH}_2\text{Cl}_2$ ):  $\lambda_{\text{max}}$  ( $A_{\text{rel}}$ ) = 704 (1.1), 643 (0.29), 574 (0.18), 534 (0.12), 453 (1), 433 (0.96), 385 (0.93), 351 (0.75) nm; Flu ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 450 nm):  $\lambda_{\text{em}}$  = 712 nm;  $\Phi_{\text{Flu}}$  ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 450 nm) = 25%. IR ( $\text{CH}_2\text{Cl}_2$ ):  $\tilde{\nu}$  = 2220 ( $\text{C}\equiv\text{N}$ ), 1733 ( $17^2\text{-C=O}$ )  $\text{cm}^{-1}$ . <sup>1</sup>H NMR ( $\text{CDCl}_3$ ):  $\delta$  = 9.15 (s, 1 H, 5-H), 9.00 (s, 1 H, 10-H), 8.37 (s, 1 H, 20-H), 7.89 (dd,  $J$  = 18, 12 Hz, 1 H, 3-CH), 6.26 (dd,  $J$  = 18, 1 Hz, 1 H, 3<sup>1</sup>-CH *trans* to 3-C-H), 6.16 (dd,  $J$  = 12, 1 Hz, 1 H, 3<sup>1</sup>-CH *cis* to 3-C-H), 5.37, 5.26 (d,  $J$  = 20 Hz, each 1 H, 13<sup>1</sup>-CH<sub>2</sub>), 4.33 (dq,  $J$  = 2, 7 Hz, 1 H, 18-H), 4.10–4.07 (br-d, 1 H, 17-H), 3.65 (s, 3 H, 17<sup>2</sup>-CO<sub>2</sub>CH<sub>3</sub>), 3.53 (q,  $J$  = 8 Hz, 2 H, 8-CH<sub>2</sub>), 3.38 (s, 3 H, 12-CH<sub>3</sub>), 3.33 (s, 3 H, 2-CH<sub>3</sub>), 3.12 (s, 3 H, 7-CH<sub>3</sub>), 2.58–2.52, 2.51–2.46, 2.29–2.24, 2.12–2.05 (m, each 1 H, 17-CH<sub>2</sub>CH<sub>2</sub>), 1.75 (d,  $J$  = 7 Hz, 3 H, 18-CH<sub>3</sub>), 1.59 (t,  $J$  = 8 Hz, 3 H, 8<sup>1</sup>-CH<sub>3</sub>), 1.03, –1.03 (s, each 1 H,  $\text{NH}\times 2$ ) ppm. MS (FAB): calcd. for  $\text{C}_{37}\text{H}_{36}\text{N}_6\text{O}_2$  596.2900; found 596.2898 [ $\text{M}$ ]<sup>+</sup>.

**Methyl 13<sup>1</sup>-Deoxo-13<sup>1</sup>-(dicyanomethylene)pyropheophorbide-*d* (**8**):** According to the reported procedure,<sup>[7]</sup> 3-vinylchlorin **7** (50 mg, 0.084 mmol) was oxidized by  $\text{OsO}_4$  (ca. 50 mg),  $\text{NaIO}_4$  (300 mg, 1.4 mmol), and AcOH (1 mL) in water (2 mL) and THF (20 mL). The crude product was purified by silica-gel chromatography ( $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$ , 1:99) followed by recrystallization from  $\text{CH}_2\text{Cl}_2/\text{MeOH}$  to give **8** (35 mg, 70%) as green crystals; m.p. 239–242 °C. UV/Vis ( $\text{CH}_2\text{Cl}_2$ ):  $\lambda_{\text{max}}$  ( $A_{\text{rel}}$ ) = 727 (1.3), 663 (0.22), 584 (0.23), 543 (0.12), 467 (0.77), 437 (0.73), 395 (1) nm; Flu ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 470 nm):  $\lambda_{\text{em}}$  = 733 nm;  $\Phi_{\text{Flu}}$  ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 470 nm) = 21%. IR (Film):  $\tilde{\nu}$  = 2222 ( $\text{C}\equiv\text{N}$ ), 1744 ( $17^2\text{-C=O}$ ), 1668 ( $3\text{-C=O}$ )  $\text{cm}^{-1}$ . <sup>1</sup>H NMR ( $\text{CDCl}_3$ ):  $\delta$  = 11.46 (s, 1 H, 3-CH), 10.17 (s, 1 H, 5-H), 9.46 (s, 1 H, 10-H), 8.69 (s, 1 H, 20-H), 5.78, 5.66 (d,  $J$  = 21 Hz, each 1 H, 13<sup>1</sup>-CH<sub>2</sub>), 4.49 (dq,  $J$  = 2, 8 Hz, 1 H, 18-H), 4.33–4.30 (br-d, 1 H, 17-H), 3.87 (s, 3 H, 12-CH<sub>3</sub>), 3.71 (s, 3 H, 2-CH<sub>3</sub>), 3.67 (q,  $J$  = 8 Hz, 2 H, 8-CH<sub>2</sub>), 3.63 (s, 3 H, 17<sup>2</sup>-CO<sub>2</sub>CH<sub>3</sub>), 3.26 (s, 3 H, 7-CH<sub>3</sub>), 2.70–2.64, 2.63–2.59, 2.34–2.29, 2.27–2.23 (m, each 1 H, 17-CH<sub>2</sub>CH<sub>2</sub>), 1.81 (d,  $J$  = 8 Hz, 3 H, 18-CH<sub>3</sub>), 1.69 (t,  $J$  = 8 Hz, 3 H, 8<sup>1</sup>-CH<sub>3</sub>), 0.75, –1.34 (s, each 1 H,  $\text{NH}\times 2$ ) ppm. MS (FAB): calcd. for  $\text{C}_{36}\text{H}_{34}\text{N}_6\text{O}_3$  598.2692; found 598.2732 [ $\text{M}$ ]<sup>+</sup>.

**Methyl 3<sup>2</sup>,3<sup>2</sup>-Dicyano-13<sup>1</sup>-deoxo-13<sup>1</sup>-(dicyanomethylene)pyropheophorbide-*a* (**9**):** Reaction of **2** (35 mg, 0.06 mmol) with malononitrile (515 mg, 7.8 mmol) in refluxing THF in the presence of  $\text{Et}_3\text{N}$  for 3.5 h gave **9** in 7% yield, whereas a similar treatment of **3** (60 mg, 0.10 mmol) with malononitrile (515 mg, 7.8 mmol) gave **9** in 12% yield. Reaction of **8** (30 mg, 0.05 mmol) with malononitrile (33 mg, 0.50 mmol) in the presence of  $\text{Et}_3\text{N}$  (35 mg, 0.35 mmol) in



THF (20 mL) at room temperature for 2 h gave **9** as green crystals in 41% yield after purification by silica-gel chromatography ( $\text{CH}_2\text{Cl}_2$ ) and recrystallization ( $\text{CH}_2\text{Cl}_2/\text{MeOH}$ ); m.p. 138–143 °C. UV/Vis ( $\text{CH}_2\text{Cl}_2$ ):  $\lambda_{\text{max}}$  ( $A_{\text{rel}}$ ) = 736 (0.97), 669 (0.22), 590 (0.21), 545 (0.13), 422 (1), 353 (0.83) nm; Flu ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 450 nm):  $\lambda_{\text{em}}$  = 749 nm;  $\Phi_{\text{Flu}}$  ( $\text{CH}_2\text{Cl}_2$ :  $\lambda_{\text{ex}}$  = 450 nm) = 20%. IR ( $\text{CH}_2\text{Cl}_2$ ):  $\tilde{\nu}$  = 2228, 2216 ( $\text{C}\equiv\text{N}$ ), 1743 ( $17^2\text{-C=O}$ )  $\text{cm}^{-1}$ .  $^1\text{H}$  NMR ( $\text{CDCl}_3$ ):  $\delta$  = 9.29, 9.17, 9.05, 8.66 (s, each 1 H, 5-, 10-, 20-H, 3-CH), 5.56, 5.39 (d,  $J$  = 20 Hz, each 1 H,  $13^1\text{-CH}_2$ ), 4.48 (dq,  $J$  = 2, 8 Hz, 1 H, 18-H), 4.28–4.25 (br-d, 1 H, 17-H), 3.66 (q,  $J$  = 8 Hz, 2 H, 8- $\text{CH}_2$ ), 3.65, 3.51, 3.46, 3.25 (s, each 3 H, 2-, 7-, 12- $\text{CH}_3$ ,  $17^2\text{-CO}_2\text{CH}_3$ ), 2.65–2.59, 2.35–2.29, 2.22–2.17 (m, 2 H, 1 H, 1 H, 17- $\text{CH}_2\text{CH}_2$ ), 1.84 (d,  $J$  = 8 Hz, 3 H, 18- $\text{CH}_3$ ), 1.67 (t,  $J$  = 8 Hz, 3 H,  $8^1\text{-CH}_3$ ), 0.47, –1.36 (s, each 1 H,  $\text{NH}\times 2$ ) ppm. MS (FAB): calcd. for  $\text{C}_{39}\text{H}_{35}\text{N}_8\text{O}_2$  647.2883; found 647.2918 [ $\text{M} + \text{H}$ ] $^+$ .

**Supporting Information** (see also the footnote on the first page of this article):  $^1\text{H}$  NMR spectra of **6–9** in  $\text{CDCl}_3$  (Figures S1–4) and relation of  $Q_y$  energy levels with redox potential differences (Figure S5).

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- [1] a) H. Scheer, in: *Light-Harvesting Antennas in Photosynthesis* (Eds.: B. R. Green, W. W. Parson), Kluwer Academic Publishers, Dordrecht, The Netherlands, **2003**, chapter 2, pp. 29–81;

- b) H. Tamiaki, R. Shibata, T. Mizoguchi, *Photochem. Photobiol.* **2007**, *83*, 152–162.
- [2] T. Mizoguchi, A. Shoji, M. Kunieda, H. Miyashita, T. Tsuchiya, M. Mimuro, H. Tamiaki, *Photochem. Photobiol. Sci.* **2006**, *5*, 291–299.
- [3] T. Mizoguchi, Y. Saga, H. Tamiaki, *Photochem. Photobiol. Sci.* **2002**, *1*, 780–787.
- [4] H. Tamiaki, M. Kouraba, *Tetrahedron* **1997**, *53*, 10677–10688.
- [5] S. Sasaki, H. Tamiaki, *J. Org. Chem.* **2006**, *71*, 2648–2654.
- [6] a) S. Sasaki, Y. Kotegawa, H. Tamiaki, *Tetrahedron Lett.* **2006**, *47*, 4849–4852; b) S. Sasaki, Y. Kotegawa, H. Tamiaki, *Bull. Chem. Soc. Jpn.* **2009**, *82*, 267–271; c) S. Sasaki, Y. Kotegawa, K. Azuma, M. Kunieda, H. Tamiaki, *Chem. Lett.* **2009**, *38*, 796–797.
- [7] H. Tamiaki, M. Amakawa, Y. Shimono, R. Tanikaga, A. R. Holzwarth, K. Schaffner, *Photochem. Photobiol.* **1996**, *63*, 92–99.
- [8] M. Katterle, A. R. Holzwarth, A. Jesorka, *Eur. J. Org. Chem.* **2006**, *2*, 414–422.
- [9] G. P. Wiederrecht, W. A. Svec, M. P. Niemczyk, M. R. Wasieleski, *J. Phys. Chem.* **1995**, *99*, 8918–8926.
- [10] A recent example of CV data of chlorophyll derivatives: C. Liu, M. P. Dobhal, M. Ethirajan, J. R. Missert, R. K. Pandey, S. Balasubramanian, D. K. Sukumaran, M. Zhang, K. M. Kadish, K. Ohkubo, S. Fukuzumi, *J. Am. Chem. Soc.* **2008**, *130*, 14311–14323.
- [11] a) X.-F. Wang, Y. Koyama, H. Nagae, Y. Wada, S. Sasaki, H. Tamiaki, *J. Phys. Chem. C* **2008**, *112*, 4418–4426; b) X.-F. Wang, O. Kitao, H. Zhou, H. Tamiaki, S. Sasaki, *Chem. Commun.* **2009**, 1523–1525; c) X.-F. Wang, O. Kitao, H. Zhou, H. Tamiaki, S. Sasaki, *J. Phys. Chem. C* **2009**, *113*, 7954–7961; d) X.-F. Wang, H. Tamiaki, *Energy Environ. Sci.* **2010**, *3*, 94–106; e) X.-F. Wang, H. Tamiaki, L. Wang, N. Tamai, O. Kitao, H. Zhou, S. Sasaki, *Langmuir* **2010**, *26*, 6320–6327.
- [12] P. H. Hynninen, in: *Chlorophylls* (Ed.: H. Scheer), CRC Press, Boca Raton FL, **1991**, pp. 145–209.

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